

Galectin-1 Regulates p53 and Is Implicated in Glioma Cell Resistance to Cytotoxic Drugs

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Galectin-1 is a potent modulator of GBM cell migration and a close partner of Ras whose importance as a signaling molecule in the case of GBMs has already been highlighted → Glioblastomas (GBM) are the most common type of primary malignant brain tumor. → Patients have an average life expectancy of one year on the basis of the standard treatment of surgical section followed by radiotherapy ➔ Galectins are differentially expressed in supratentorial pilocytic → GBM are associated with dismal prognoses because they diffusely infiltrate the brain parenchyma, and manna astrocytomas, astrocytomas, anaplastic astrocytomas and glioblastomas because migrating malignant glioma cells are resistant to apoptosis, and thus to pro-apoptotic cytotoxic drugs and significantly modulate tumor astrocyte migration. due to constitutive activation of distinct anti-apoptotic signaling pathways including: PI3-K, Akt/PKB Camby et al., Brain Pathol, 2001 (PTEN), mTOR, NF-kappaB, etc → Galectin-1 modulates human glioblastoma cell migration into the brain through modifications to the actin cytoskeleton and levels of expression of small GTPases. Camby et al., J Neuropathol Exp Neurol, 2002 → Galectin-1: A small protein with major functions. Camby et al., Glycobiology 2006, in press. We investigated whether decreasing the levels of expression of galectin-1 in human Hs683 GBM cells could increase their sensitivity to the pro-autophagic effects of temozolomide. An anti-galectin-1 siRNA approach was employed to decrease the expression levels of galectin-1 in human Hs683 GBM cells Temozolomide displays actual efficacy against malignant gliomas (Stupp et al., N Engl J Med 2005) because it is a pro-autophagic drug, not a pro-apoptotic one (Kanzawa et al., Cell Death Differ 2004) Temozolomide Increases Galectin-1 Expression in Human Hs683 GBM Cells In Vitro and In Vivo Designing Anti-Galectin-1 siRNAs Vestern blot analyses of Gal1 expression in Hs683 GBM cells left eated (CT) or treated with 10µM and 100µM temozolomide for Autoritation (CT) or treated with TopM and 100pM temocolomide for Th. Th. B: Immunfluxrescence staining to reveal galectin -1 expression (green funcrescence) in with type H583 cells (ell nutrested (Ba) or treated with 10pM temozohemisc for 72h (Bb). C: In vivo expression of Gall detected by immundhistchemistry in the GIBM of H583 cell-barring immuno-compromized mice left untrested (Ca) or treated with temozohomide (40mg kg is / three times a week for three weeks; Cb). Dar: Western block533. Dbs Sarviva of Immuno-compromized mice fail expression 5 to 9 days post transfection with anti-Gall stRNA (is see Fig.) Of 16853. Dbs Sarviva of Immuno-compromized mice after frain grafts of i) wild type H5832 cells (blue line), ii) H5832 cells attrasteted with temozohomide (Temodal; hatched lines) at 40 mg/kg ver treated with temozohomide (Temodal; hatched lines) at 40 mg/kg very torther unes a week (ack Monday). Wothavday and Friday) for three consecutive weeks, with treatment starting on the 5th day root-tumer gaft. ccession number NM_002305, 526 bp) with indication of putative siRNA sequences: si-1 (first underlined b-14 Α Da 10uM 100uM ст unce below), si-2 (second underlined sequence below) and si-3 (third derlined bold sequence below). acagcaacaa cotgtgoot Db tcaaget gecagatgga taegaatt Cumulative Prop Surviving 1 on of Gal1 (green fluorescence) detected by zence in Hs683 GBM cells transfected for 5 days with si-1, si-2 si-3, scrambled siRNA (scr) or with calcium phosphate transfection buffer (Ca) compared to that of wild-type (wt) Hs683 cells. B: Expression of Gal1, Gal2, Gal3 and Gal9 detected by immunofluorescence Hs683 GBM cells transfected for 5 days with si-1 anti-Gal1 siRNA or with th day post-tumor graft Decreasing Galectin-1 Expression in Hs683 GBM cells Increases the Survival of Hs683 GBM Orthotopic Xenograft-Bearing Immuno-Compromized Mice and Improves the Therapeutic Benefits of Temozolomide Decreasing Galectin-1 Expression in Hs683 GBM cells Increases the In Vitro Anti-Tumor Effects of cytotoxic Drugs Db Ca day 5 day 7 day 9 LC3-I Wound Healing 30 LC3-II Cb wt scr si wt scr si wt scr si Percentages 10 116 kDa scr Duration of Wound Healing A: Quantitative determination (computer-assisted videomicroscopy) of the ability of Hs683 GBM cells to Da: Flow cytometry analyses of acridine orange staining measuring the red fluorescence in wild type (wt), semabled iRNA- (we) and anti-Ga1 iRRNA (a) transfected H6683 GBM cells treated for 72h with 10µM temosolimic (black ktrus) or lef untreated (open hars). Due: LC3 western blotting analyses of 1) wt, ser or anti-Ga11 (a) transfected H6683 cells. E: Flow cytometry analyses of acriding the compass transmission green fluorescence in wild type (wt), semabled iRRNA- (wr) and anti-Ga11 iRNA (a) transfected H6683 GBM cells treated for 72h with 10µM temosine (solve) and anti-Ga11 iRNA (a) transfected H6683 GBM cells treated for 72h C: Apoptosis measurements by means of flow cytometry analyses by TUNEL (Ca) or PARP clavage analyses (Cb) in wt, scr and anti-Gall transfected Hs683 cells (si). The open bars in Ca represent Hs683 cells left untreated, while black bars represent cells treated for 72h with 10µM colonize the "mechanical" wound made at 0 hour on a cell population grown to confluence (Scratch wound assay). Open bars: cells were transfected with scrambled siRNA and left untreated. Blue bars: cells were transfected with scrambled siRNA and treated with 10µM temozolomide. Black bars: cells were transfected temozolomide. with anti-Gal1 siRNA and left untreated. Grey bars: cells were transfected with anti-Gal1 siRNA and treated rith 10µM tem B: Percentages of killed cells assessed by the colorimetric MTT assay in Hs683 GBM cells: left unt with 10µM temozolomide (black bars) or left untreated (open bars).
F: Immunofluorescence analyses of cathepsin B expression and localization in scr (Fa) and anti-Gall (open bars), transfected with scrambled siRNA (grey bars) or transfected with the anti-Gall siRNA (black bars) and treated with BCNU, PCB, CCNU (10µM) or VCR (10nM) for 72h. siRNA (si: Fb) transfected Hs683 cells. Decreasing Galectin-1 Expression in Hs683 GBM cells Does Not Induce Apoptosis, Autophagy or LMP CONCLUSION Decreasing Galectin-1 Expression in Hs683 GBM cells Does Not Impair Classical Pathways Involved in resistance to Chemotherapy but Impairs the Endoplasmic Reticulum Stress (ERS) Response → Temozolomide treatment increases galectin-1

A: Western blots illustrating the expression and phosphorylation levels of Aki in Hs683 cells that have been i) left untreated (wt) or ii) transfected with scrambled siRNA (ser) or iii) transfected with anti-Gall siRNA (si) or antigalectin-3 siRNA.

B: Illustration of the effects of Gall depletion (si; see Fig. 1) as compared to controls (wt and ser Hs683 cells) on the expression level (open bras in **Ba** and **Bb**) or the level of phosphorylation (black bars in **Ba** and **Bb**) of Src (**Ba**), PJSL-pS8 (**Bb**) and MF-sB activation (pd5/ReL1 0MA binding activity) (**Bc**).



C: Immunofluorescence analyses (with bright field controls) of DUSP5 expression in set (Ca-Cb) and anti-Gall (Ca-Cd) aRNA transfected H683 cells. De Westen blois Milarating the expression levels of ATF3 in wt, set and anti-Gall siRNA (si) transfered H683 cells. E: Immunofluorescence analyses (with bright field controls) of HERP expression in set (Ea-Eb) and anti-Gall (Er-Ed) siRNA transfered H683 cells. Fb) and anti-Gall (Fe-Fd) siRNA transfered H683 cells.

- → Temozolomide treatment increases galectin-1 expression
- ➔ Decreasing galectin-1 expression in glioma cells increases the anti-tumor effects of chemotherapeutical agents
- → Decreasing galectin-1 expression does not induce apoptotic or autophagic features
- → Decreasing galectin-1 expression weaken glioma cell defenses by impairing their ERS response
 - Taken together, these effects seem to reinforce the therapeutic benefit of temozolomide in *in vivo* glioblastoma models.
- The novel aspects of galectin-1-related function in the ERS response may be amenable to therapeutic manipulation either by the *in vivo* delivery of antigalectin-1 siRNA as demonstrated here, or through compounds suppressing galectin-1.